Archives of Clinical Microbiology 1989-8436 2023

Vol. 14 No. 3: 232

Human Prion Diseases Tvisha

Tvisha Joshi*

Dublin High, 8151 Village Pkwy, Dublin, CA 94568, USA

Corresponding author: Tvisha Joshi

nidhi_ahuja@berkeley.edu

Dublin High, 8151 Village Pkwy, Dublin, CA 94568, USA

Citation: Joshi T (2023) Human Prion Diseases. Arch Clinic Microbio, Vol. 14 No. 3: 232.

Abstract

The functions of these normal prion proteins are still not completely understood. The discovery of proteinaceous infectious agents by Prusiner in 1982 was sensational. All previously known pathogens contained nucleic acids, the code of life, that enabled them to reproduce. In contrast, the proteinaceous agents of disease, called prion proteins (PrP), lacked nucleic acids and propagated by binding to the functional, endogenous form of cellular prion protein (referred to as PrP^c) and altering its conformation to produce the infectious disease-causing misfolded protein (referred to as PrP^{sc}). The accumulation and aggregation of these infectious prion proteins within the brain causes destruction of neural tissue and leads to fatal spongiform encephalopathies. In this review, we present the molecular pathology of prion-based diseases. These insights are of particular importance since the principles of prion pathogenesis apply to other neurodegenerative disease such as Alzheimer's disease, Huntington's disease, Parkinson's disease and amyotrophic lateral sclerosis. Collectively, the global prevalence of these diseases is rapidly increasing while effective therapies against them are still lacking. Thus, the need to understand their etiology and pathogenesis is urgent, and it holds profound implications for societal health.

Keywords: Neurodegeneration; Pathology; Plaque; CJD; Scrapie

Received: 21-Feb-2022, Manuscript No. ipacm-23-13508; **Editor assigned:** 24-Feb-2023, Pre-QC No. ipacm-23-13508 (PQ); **Reviewed:** 01-Mar-2023, QC No. ipacm-23-13508; **Revised:** 14-Mar-2023, Manuscript No. ipacm-23-13508 (R); **Published:** 24-Mar-2023, DOI: 10.36648/1989-8436X.22.14.03.232

Historical Background

For nearly half a century, baffled physicians had watched their patients suffer from a rapidly progressive deterioration of brain functions [1, 2]. This neurodegenerative disorder that ultimately killed the patients was named as Creutzfeldt-Jakob disease (CJD). It was observed that while CJD destroyed the brain of patients and dementia took over, the immune system was slow to respond. In 1959, Igor Klatzo, Carleton Gajdusek, and Vincent Zigas recognized the similarities between the pathologies of CJD and another neurodegenerative disease called kuru that was prevalent in natives of New [3]. In that same year, William Hadlow noticed how signs of kuru were similar to those of scrapie [4]. It was believed at the time that scrapie was caused by a "slow virus", as proposed by Bjorn Sigurdsson in 1954 while he was conducting studies on scrapie sheep in Iceland [5]. The transmissibility of kuru and CJD was established by Gajdusek, Clarence Gibbs, and their colleagues, who had passaged these diseases to chimpanzees (Gajdusek et al., 1966). Scrapie had been previously shown to be transmissible in sheep [6]. For many years, it was incorrectly assumed that these three neurodegenerative diseases, namely

CJD, kuru and scrapie, resulted from infections of slow or latent viruses but we now know that all of these diseases are caused by prions.

Prion Diseases In Humans

Creutzfeldt Jakob disease

In humans, CJD has been the most studied prion-based disease. It is a rapidly progressing neurodegenerative disorder that affects approximately one person in a population of a million every year. In the United States of America, this statistic equates to roughly 330 new cases in a year, and among these, about 85% cases are of sporadic CJD that occur without a discernible environmental source of infection. Sporadic CJD causes progressive neurologic dysfunction in multiple areas of the brain that may be associated with alteration of personality or behavior, insomnia, anorexia, and depression, involuntary movements, severe cognitive impairment, and death within a few months [7]. In about 5% of patients with sporadic CJD, the onset of neurologic dysfunction is so abrupt that may be confused with a stroke. Familial CJD, unlike sporadic CJD, results from mutations of the gene for PrP, and accounts for up to 15% cases. The features of familial CJD vary according to the underlying mutation, but in general, there is an earlier age of onset in familial cases along with a more prolonged duration of illness. Contaminated medical instruments or biological material such as corneal grafts, human dura mater grafts, and human pituitary-derived hormones have been documented to result in a few cases of acquired CJD known as iatrogenic CJD. Signs and symptoms of iatrogenic CJD depend on the route of inoculation. When the route of inoculation is through the central nervous system then the clinical presentation of the disease is like that of sporadic CJD, whereas with a peripheral route of inoculation an ataxic syndrome results. In the late 1980s, a variant form of acquired CJD (vCJD) was identified in people who had eaten contaminated beef that was derived from cows suffering from bovine spongiform encephalopathy. In vCJD, the cases initially present psychiatric symptoms such as depression and withdrawal for months prior to the development of neurologic signs [8]. The terminal stages of vCJD appear similar to those of sporadic CJD.

Kuru

The word Kuru in fore means "to shake" and it is used to describe a slow progressing fatal infection of the brain. It is a transmissible spongiform encephalopathy that is associated with deposits of kuru amyloid plaques in the cerebellum, thalamus, and cerebral cortex that are followed by severe gliosis of astrocytes, neuronal lesions, and spongiform changes within the brain. Kuru became epidemic in New Guinea between 1950-1970 because of the endo-cannibalistic funeral rituals practiced by the native Fore population. The investigations of the Fore people by anthropologists suggested that kuru had begun in the region between 1900 and 1920 [9]. While the incubation period for kuru can be several decades, death usually occurs within months of the onset of symptoms. Initially, Kuru presents with progressive trembling, an unsteady gait and dysarthria. With time, muscle tremors become more severe, ataxia worsens, and the patient loses voluntary muscle control. Unlike other prion diseases, severe dementia does not occur in kuru [10]. Currently, this disease is considered nearly eradicated due to the elimination of tribal cannibalism.

Gerstmann-Sträussler-Scheinker (GSS) disease

Alterations in the PrP gene sequence can result in heritable prion-based diseases. Virtually all cases of GSS result from such inherited mutations of the PrP gene. Rarely, a mutation may appear spontaneously in the PrP gene in individuals without a family history of the disease. These are called de novo variants that are associated with gene mutation appearing at the time of the formation of the egg or sperm for that individual only, with other family members not affected. The de novo variant can be passed on to the next generation in an autosomal dominant manner. Those who possess a disease-causing mutation in PrP gene can eventually develop signs and symptoms because of neurodegeneration, however the progression and the overall severity of their disorder can vary depending upon on the underlying mutation. A common symptom that appears early is a progressive loss of coordination that may be seen as unsteady gait, difficulty walking, and inept movement of limbs. As the disease progresses, other symptoms start appearing such as problems associated with cognition, language, memory, and behavior. In GSS, PrP^{sc} can be found in many parts of the brain, including cerebellum and thalamus, and this isoform can transmit the disease to experimental animals [11].

Fatal Familial Insomnia (FFI)

The D178N mutation of PrP gene is associated with the development of a progressive sleep disorder called FFI. In patients who have the PrP D178N mutation along with a methionine at the 129th position of PrP, fatal sleep disorder presents itself after the age of 50 and death occurs within a year of onset of symptoms. In the brains of these patients, deposits of PrPsc plaques are confined mainly to the thalamus. In contrast, when the PrP D178N mutation is combined with a valine at 129th position of PrP, then it results in development of familial CJD. These patients experience dementia, and deposits of PrP^{sc} plaques can be seen throughout the brain upon postmortem. Clinical signs of FFI include altered sleep-wake cycles with decreased vigilance, dysfunction of the autonomic system that presents as systemic hypertension and irregular breathing, and motor manifestations such as diplopia, dysphagia, dysarthria, ataxia and abasia. Sleepwake and vigilance abnormalities are characterized by insomnia, hallucinations, and confusion. The clinical signs can vary according to the mutation at codon 129.

PrP biology

The gene encoding for PrP is present on the human chromosome #20. After its transcription, the translation of the entire reading frame occurs from a single RNA exon. This removes the possibility that PrP^{sc} plaque production or development of prion diseases results from alternative splicing of its RNA. During development, the PrP mRNA expression is stringently regulated, but in adults, this mRNA is constitutively expressed with its highest levels being produced in the neurons. Upon translation, the mRNA produces a polypeptide that is 253 amino acids long and has several key features including an amino-terminal signal peptide, a conserved hydrophobic region in the center followed by another hydrophobic region at the carboxy-terminus that serves as an attachment site for a glycosylphosphatidylinositol (GPI) anchor. Another interesting feature of this polypeptide is its repeat sequences that are comprised of one copy of the nonapeptide PQGGGGWGQ and four copies of the octapeptide PHGGGWGQ. Majority of PrP^c are found attached to the outer face of the cell membrane by their C-terminal GPI anchors. The pathway followed by PrP^c for this targeting is same as that of other membrane-bound proteins which involves synthesis on the rough ER and sorting through Golgi before reaching the cell surface. Along this path, PrP^c is post translationally processed to allow for the removal of a signal peptide from the amino terminus and of a stretch of 23 amino acids from the carboxy-terminal, a disulfide bridge formation, the addition of oligosaccharides at the amino terminus and of a GPIanchor at the carboxy-terminus. The processed protein is ~208 amino acid long chain that folds into two domains of roughly equal size. Studies have demonstrated that the attachment of a GPI anchor along with the number of oligosaccharides attached to PrP can vary. Many different cell types, including neurons, secrete

PrP^c without its anchor to such an extent that it can accumulate to 10% of total PrP^c in brain extracts. The absence of GPI anchor does not automatically cause PrP misfolding, but it has been observed that PrP anchored on the cell surface is important in controlling neuronal toxicity. Anchored PrP^c does not stay on the cell surface long after its delivery there, but rather it recycles between the cell membrane and an endocytic vesicle.

Prion protein is highly conserved among mammals, so it is surprising that animals lacking PrP^c develop and behave normally. Interestingly, animals that don't express prion proteins do not succumb to prion disease. Several cellular functions have been proposed for PrP^c. These include regulation of myelination, copper metabolism, cell communication, cell migration, cell differentiation, cell survival, circadian rhythm, and inhibition of Bax mediated apoptosis.

Prion propagation

A small number of PrP^{sc} presented to the host cell can corrupt endogenous PrP^c and convert them all to PrP^{sc}. During this transition, the shift in secondary structure of the protein becomes obvious: α -helices predominate the secondary structure of PrP^c, whereas β -sheets predominate in PrP^{sc}. This spontaneous α -to- β transition in the structure of prion protein is the most fundamental event that defines the pathogenesis of prion diseases. This structural change is associated not only with loss of function but with aggregation and pathological amyloid assembly. The initial trigger for amyloid assembly may not always be exogenous PrP^{sc}. Mutations that destabilize the structure of endogenous prion protein can promote amyloid formation and leads to prion diseases with sporadic or genetic origin.

The molecular mechanism that converts PrP^c to PrP^{sc} is not conclusively known. However, several models have been proposed, including a direct conversion mechanism, whereby each protein in the PrP^{sc} state can bind to PrP^c to form a heterodimer and then through this interaction convert PrP^c to PrP^{sc}. Another model suggests that a cofactor may unfold PrP^c to a partially unfolded PrP* state that may facilitate formation of the heterodimer with PrP^{sc} (Cohen et al., 1994). Either way, subsequent addition of PrP^c to the heterodimer results in fibril formation, its elongation that ultimately leads to amyloid assembly. Depending on its conformational stability, the growing fibril may break and release new templates that spread and continue to support the chain reaction of prion protein misfolding and aggregation in different parts of the brain.

Although, the intracellular location of PrP^{Sc} formation is not known yet but the conversion of the PrP^C to PrP^{Sc} occurs in caveolae-like domains. After this conversion, PrP^{Sc} starts accumulating in the lysosomes and in the late endocytic vesicles of the affected cells. This intracellular accumulation of PrP^{Sc} induces vacuolation of neurons and hypertrophy of reactive astrocytes that eventually kills the neurons. At this point, PrP^{Sc} begin to accumulate

extracellularly. PrP^{sc} polymerize to form amyloid plaques that leads to more death of neurons, progressive destabilization of neuronal networks and spongiform degeneration of the brain that is a hallmark of prion diseases.

Prion-like neurodegenerative diseases

The clinical presentation of human prion diseases bears striking resemblance to the pathological hallmarks of many other commonly known progressive neurodegenerative disorders. The common signs include memory loss, dementia, difficulties with movement, loss of brain function, and changes in behavior and personality. At a molecular level, these neurodegenerative diseases are characterized by accumulation of misfolded protein deposits that correlate with progressive loss of neurons and accumulation of amyloid lesions in the brain. The multimeric amyloid assemblies of misfolded proteins implicated in these diseases include amyloid- β plaques in Alzheimer's disease, α -synuclein Lewy bodies and tau neurofibrillary tangles in Parkinson's disease, huntingtin protein aggregates in Hungtington's disease, and multiple protein aggregates in Amyotrophic Lateral Sclerosis. The endogenous proteins implicated in each of these diseases use similar mechanisms for misfolding, polymerization and propagation. A common theme seen in these proteins is that they all misfold to conformations that are rich in β -sheets. In all these cases, the misfolded proteins lead to chain reactions whereby they act as template to corrupt the conformation of normal proteins. These misfolded proteins aggregate to form amyloids that disrupt function either by directly harming nearby cells, and/or by sequestering essential proteins that are unable to execute their role within the body. The ability of these proteins to propagate by the process of conversion followed by aggregation, allows the disease to be initiated by a small number of misfolded proteins that set-in motion a rapidly progressing disorder. Since the normal and misfolded isoforms of the proteins have the same sequence and since the host is already tolerant to the normal isoform so these diseases do not elicit a severe immune response.

Conclusion

It has been 40 years since *pr*oteinaceous *in*fectious particle were discovered. They have been studied a lot since then but the mechanisms by which they cause neurodegeneration have not been fully uncovered. Pathology of other neurodegenerative diseases where misfolded aggregated proteins are implicated is also poorly understood. Understanding how PrP^c misfolds into PrP^{sc} and how these corrupted proteins form toxic aggregates may open new avenues to develop effective therapies not just for prion diseases but also for other common neurodegenerative disorders too.

Conflict of Interest Statement

The authors had no conflicts of interest to declare in this article.

References

- 1 Creutzfeldt HG (1920) Über eine eigenartige herdförmige erkrankung des zentralnervensystems. Mol Aspects Med 15: 195-291.
- 2 Jakob A (1921) Über eigenartige erkrankungen des zentralnervensystems mit bemerkenswertem anatomischen befunde. Z Gesamte Neurol Psychiatr 64: 147-228.
- 3 Klatzo I, Gajdusek DC, Zigas V (1959) Pathology of kuru. Laboratory Investigation 8: 799-847.
- 4 Hadlow WJ (1959) scrapie and kuru. Lancet 2: 289-290.
- 5 Sigurdsson B (1954) RIDA chronic encephalitis of sheep: With general remarks on infections which develop slowly and some of their special characteristics. British Veterinary Journal 110: 341-344.
- 6 Roods R, Gajdusek DC, Gibbs CJ (1973) the clinical characteristics of transmissible Creutzfeldt-Jakob disease. Brain 96: 1-20.

- 7 Will RG, Ironside JW, Zeidler M, Cousens SN, Estibeiro K, et al. (1996) a new variant of creutzfeldt-jakob disease in the UK. The Lancet 347: 921-925.
- 8 Liberski PP, Gajos A, Sikorska B, Lindenbaum S (2019) Kuru, the first human prion disease. Viruses 11: 232.
- 9 Nonno R, Angelo Di, Bari M, Agrimi U, Pirisinu L, et al. (2016) Transmissibility of gerstmann-sträussler-scheinker syndrome in rodent models: New insights into the molecular underpinnings of prion infectivity. Prion 10: 421-433.
- 10 Goldfarb LG, Petersen RB, Tabaton M, Brown P, LeBlanc AC, et al. (1992) Fatal familial insomnia and familial creutzfeldt-jakob disease: Disease phenotype determined by a DNA polymorphism. Science, 258: 806-808.
- 11 Goldfarb L, Haltia M, Brown P, Nieto A, Kovanen J, et al. (1991) New mutation in scrapie amyloid precursor gene in finnish creutzfeldtjakob kindred. The Lancet 337: 425.